

Quick Guide for PyroMark[™]Q24 Software

Start PyroMark[™]Q24 Software

In the Windows **Start** menu, select **All Programs** | **Biotage** | **Pyromark Q24**.

Context-sensitive help can be accessed at any time by pressing the **F1** key.

Set Up an Assay

- In the shortcut browser, right-click the folder you want to place the assay file in and select New Assay and then AQ Assay or CpG Assay from the context menu.¹
- 2. Enter the file name and press the **Enter** key.
- 3. Type or paste the Sequence to Analyze.
- 4. Click the Generate Dispensation Order button.
- 5. Click 🛃 in the toolbar.²
- 6. Before running your samples, validate your assay using reference samples; see the *Assay Design and Validation* section in PyroMark[™]Q24 User Manual.

Optional

If desired, enter a **Note** about the assay and set up the variable positions at the **Variable Positions** tab.

If creating a CpG assay, we recommend that you add bisulfite treatment controls by left-clicking a bold, orange **T** or **A** in the histogram. Preferable in the beginning of the sequence.



Note: In the sequence before bisulfite treatment (can be entered in the assay), check whether the suggested bisulfite controls are Cs converted to Ts (read as Gs and As in a reverse assay) or not.

Set Up a Run

- In the shortcut browser, right-click the folder you want to place the run file in and select New Run from the context menu.¹
- 2. Enter the file name and press the Enter key.
- 3. Select **Instrument Method**; see the instructions supplied with the used reagents and cartridge.
- 4. Set up the plate:
 - a. Add an assay to each used well, e.g. drag an assay in the shortcut browser to a well or a selection of wells.

Plate Setup

_					
	1	2	3	4	
	CpG Assay 1	CpG Assay 2	AQ Assay 1	AQ Assay 2	
A					

A well is colored according to the assay loaded to the well.

b. To enter a sample ID or note, select the cell and enter the text.



A selected cell is highlighted with a blue background color.

- 4. Click 🔙 in the toolbar.²
- 5. Print a list of required volumes of reagents and the plate setup; select **Pre Run Information** from the **Tools** menu and, when the report appears, click **a**.
- 6. Close the run file and copy it to one of the USB memory sticks supplied with the system.

Optional

If desired, enter the **Reagent ID** (i.e. the lot number for PyroMark[™]Q24 Gold Reagents), a **Plate ID**, a **Barcode** number for the plate, and a **Run Note**.

- ¹ To add a shortcut to a folder in the shortcut browser, click **Add Folder Shortcut**.
- ² The more frequently a file is saved, the more information is recovered if there is a power failure or similar problem while the file is open. To secure the data, a backup of PyroMark[™]Q24 files should be performed frequently.



Analyze the Run

- 1. Move the processed run file from the USB memory stick to a computer running PyroMark[™]Q24 Software.
- 2. Open the run file by double-clicking the run file (in the shortcut browser.1
- 3. At the **Overview** tab, either analyze all wells or a selection of wells with a valid analysis setup for the selected analysis mode.



Analysis Modes

AQ assays are analyzed in the AQ mode and CpG assays are analyzed in the CpG mode. SNP genotyping can be accessed through the AQ mode. To toggle between the modes, select AQ or CpG in the toolbar.

Note: How the analysis is performed can be modified at the **Analysis Setup** tab.

View the Analysis Results

Select an analyzed well at the **Overview** tab and the following information is shown:

• Well information

Assay name, sample ID, note, and any analysis warnings are listed in the **Well Information** area.

Pyrogram[®]

The analysis results (the allele frequencies or the methylation percentages) are displayed above the variable positions in Pyrogram, for example 96%.



Variable positions are highlighted with a blue-gray background color, and bisulfite treatment controls with a light yellow background color.

Quality Assessments

The quality assessments for the variable positions are displayed by:

• Quality bars in the plate overview at the **Overview** tab.

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• The background color of the analysis results in Pyrogram.

= Not analyzed¹



= Failed



¹ Either analysis is not supported by the software (e.g. SNP when in the CpG mode) or the variable position has been deselected by the user.

Methylation Levels

When in the CpG mode, a methylation bar at the Overview tab shows the methylation level for each CpG site in the well.



= Above the defined range



Analysis Reports

To generate a report, select the desired report from the Reports menu. For more information about the reports, see the online help.

ts	W	Rep	oorts	Window	Help		
CpG Ana			AQ Analysis Statistics				
CpG Ana			AQ Analysis Results				
CpG Pyre			AQ Pyrogram Report				
CpG Full			AQ Full Report				
		SNP Analysis Results					
			SNP	Pyrogram R	eport		
			SNP	Full Report			
			SNP	Overview R	eport		

More Information

Context-sensitive help is accessed by pressing the **F1** key when in a dialog or window in the software.

Repo

United States: PyroMark[™]Q24 System is designed for Laboratory Use Only, which means it may be used for either research purposes or by high complexity CLIA certified laboratories. Europe: PyroMark 1224 System is available for research and, in certain European countries*, for in vitro diagnostic applications. PyroMark 224 System meets the requirements of Annex III of the European Directive for In Vitro Diagnostic Medical Devices 98/79/EC.

* For more information, see www.biotagebio.com.

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Quick Guide for PyroMark[™]Q24 Vacuum Prep Workstation

Detailed instructions are available in PyroMark[™]Q24 User Manual.

I. Immobilize the PCR Product to Beads

- Gently shake the bottle with Sepharose beads* from side to side until a homogenous solution is obtained.
- 2.





Beads* 2 µl/sample

Binding Buffer[†] 40 µl/sample

MIX

Add 10–20 µl of PCR product/well

Total volume = $80 \,\mu\text{L/well}$

60-70 µl/well



II. Separate the DNA Strands and Release the Samples in PyroMark[™]Q24 Plate



Turn for instructions

III. Anneal the Samples to a Sequencing Primer



Heat at 80°C for 2 minutes and cool to room temperature for at least 5 minutes.

The plate can now be processed in PyroMark[™]Q24 Instrument.

IV. When You Have Finished the Day's Work

At the end of a working day, liquid waste and any solutions left in the troughs should be discarded; see instructions in PyroMark[™]Q24 User Manual.

* Streptavidin Sepharose High Performance (34 µm, 5 ml, GE Healthcare)

[†] A product from Pyrosequencing[®]

3. Seal using strip caps.

4. Agitate constantly for

5–10 minutes, 1400 rpm.

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II. Separate the DNA Strands and Release the Samples in PyroMark[™]Q24 Plate

- 1. Fill a plate (PyroMark[™]Q24 Plate) with 0.3 µM sequencing primer in 25 µl Annealing Buffer[†] in each well.
- 2. Open the vacuum switch on the tool (ON) and wash the filter probes by flushing them with approximately 70 ml water[‡]. Use the trough to the left of Parking.
- 3. Fill the troughs with indicated volumes.



⁺ Milli-Q 18.2 M Ω x cm (Millipore Corporation) or equivalent



ON

WARNING! Irritant.

The Denaturation Solution contains sodium hydroxide. MSDS can be downloaded at www.biotagebio.com.



Quick Guide for PyroMark[™]Q24 Instrument

Start the Instrument

Turn on (I) the instrument using the power switch at the rear of the instrument. A light indicator in the left corner of the screen is lit when the instrument is turned on. The screen is blank during start-up, which may take up to one minute.





The instrument shall be connected to properly grounded mains outlets using the supplied power supplies; see the Installation and Safety document.

Load the Reagents and Plate







- Fill a reagent cartridge according to the Pre Run Information report and the 1. instructions supplied with the reagents (PyroMark[™]Q24 Gold Reagents).
- 2. When the instrument is not processing, open the instrument lid. An audible warning signal will alert you if the lid is opened when it is not safe.
- 3. Open the cartridge gate and insert the cartridge. Push it the whole way in and then down.
- Ensure that the line is visible in front of the cartridge and close the gate. 4.
- Open the plate-holding frame and place the plate on the heating block. 5.
- Close the frame and the instrument lid. 6.



due to moving parts. Keep the instrument lid closed while the instrument is processing.

WARNING! Sharp nedles at the bottom of the reagent cartridge.



60-0277 AA

Start and Monitor a Run



Run files are transferred to the instrument using the USB memory sticks supplied with the system.

- 1. Plug the USB memory stick containing the run file into the USB port.
- 2. Select **Run** in the main menu (using the \blacktriangle and \checkmark screen buttons) and press **Ok**.
- 3. Select the run file and press Select.

The instrument will start dispensing reagents when the pressure in the dispensing unit, the speed of the mixer, and the temperatures of the heating block, process chamber lid, and the coolant liquid have reached their preset levels (may take several minutes).

To show Pyrogram[®] of another well, use the ▲ and ▼ screen buttons.

When the Run is Finished

- 1. When the instrument confirms that the run has been finished, press **Close**.
- 2. Remove the USB memory stick. (If it was removed before the run was finished, insert the memory stick and select **Administration** and then **Copy Unsaved Runs**.)
- 3. Remove and discard the plate.
- 4. Remove the cartridge (by opening the gate and lifting the cartridge up and pulling it out) and discard or clean it. Refer to PyroMark[™]Q24 User Manual for cleaning instructions.
- 5. Analyze the run on an office computer running PyroMark[™]Q24 Software.

When You Have Finished the Day's Work

- 1. Shut down the instrument.
- 2. At the rear of the instrument, press the light button and check that the coolant level is visible in the window. If not, please contact 1-Point Support (see footer).
- 3. If the instrument needs to be cleaned, see instructions in PyroMark[™]Q24 User Manual.

Shut Down the Instrument

- 1. Select **Shutdown** in the main menu and press **Ok**.
- 2. When the message *It is now safe to turn off the instrument* appears, turn off (0) the instrument. The power switch is located at the rear of the instrument.









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